



## Draft Genome Sequence of *Microbacterium profundi* Shh49<sup>T</sup>, an Actinobacterium Isolated from Deep-Sea Sediment of a Polymetallic Nodule Environment

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Microbacterium profundi strain Shh49<sup>T</sup> was isolated from deep-sea sediment from a polymetallic nodule area located in the East Pacific Ocean. Strain Shh49<sup>T</sup> contains genes related to the reduction/oxidation of metals. It has potential application in the bioremediation of heavy metal-contaminated environments.

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cean polymetallic nodules are widely distributed on the deepsea floor and are a potential source of metals, such as Fe, Mn, Ni, Cu, and Co (1). Microbes participate in the formation of polymetallic nodules (2). *Microbacterium profundi* Shh49<sup>T</sup> belongs to the class *Actinobacteria*. It was isolated from deep-sea sediment of a polymetallic nodule environment (8°22′38″N 145°23′56″W) at a depth 5,280 m (3). To understand the ecological function of strain Shh49<sup>T</sup> and its potential role in the formation of nodules, the genome of strain Shh49<sup>T</sup> was sequenced and analyzed.

Genomic DNA sequencing was performed using Solexa paired-end sequencing technology (HiSeq 2000 system; Illumina, Inc., USA) (4) by a whole-genome shotgun (WGS) strategy, with a 500-bp paired-end library (333 Mb available reads, 100-fold genome coverage) and a 2,000-bp paired-end library (140 Mb available reads, 42-fold genome coverage). All clean reads were assembled using SOAP*denovo* version 1.05 (5, 6). The quality of the sequencing read data was estimated by G+C content and sequencing depth correlation analysis. The tRNAs and rRNAs were identified using tRNAscan-SE (7), RNAmmer (8), and the Rfam database (9). The open reading frames (ORFs) and the functional annotation of translated ORFs were predicted and achieved using the RAST server online (10). The classification of some predicted genes and pathways was analyzed using the KEGG databases (11–13).

The draft genome sequence of strain Shh49<sup>T</sup> revealed a genome size of 3,369,357 bp (scaffold length) and was assembled into 12 contigs. The G+C content was 66.54%. These scaffolds contain 3,269 coding sequences (CDSs), 44 tRNAs, and one copy of 16S-23S-5S rRNA gene operons. Among the protein-coding genes, 2,355 were assigned to putative functions, and the remaining were annotated as hypothetical proteins.

To study the ecological function of strain Shh49<sup>T</sup> in the metal cycle, the reductase/oxidase relationship to metal reduction/oxidation, including Fe, Mn, Cu, and Hg, was analyzed. Four multicopper oxidases (MCOs), a family of enzymes known to be in-

volved in Fe (14), Cu (15, 16), and Mn oxidation (17), were detected. Strain Shh49<sup>T</sup> may have potential ability to oxidize iron from ferrous to ferric iron on the basis of the detection of two ferroxidases. Further experiments need to be performed to confirm its function. Strain Shh49<sup>T</sup> encodes one mercuric ion reductase and one flavin adenine dinucleotide (FAD)-dependent NAD(P)-disulfide oxidoreductase, both of which participate in the reduction of Hg<sup>2+</sup> to Hg. The genome of strain Shh49<sup>T</sup> may further help us investigate the cycles of metals in deep-sea polymetallic nodules. Strain Shh49<sup>T</sup> also has potential application in the bioremediation of heavy metal-contaminated environments.

Nucleotide sequence accession number. The draft genome sequence of strain Shh49<sup>T</sup> is available in GenBank under the accession no. JPSY00000000.

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## **REFERENCES**

- Banakar V, Nair R, Parthiban G, Pattan J. 2000. Oceanic ferromanganese deposits: future resources and past-ocean recorders. Indian Miner 34:41–56.
- Wang X, Schlossmacher U, Wiens M, Schröder HC, Müller WE. 2009. Biogenic origin of polymetallic nodules from the clarion-Clipperton Zone in the eastern Pacific Ocean: electron microscopic and EDX evidence. Mar Biotechnol 11:99–108. http://dx.doi.org/10.1007/s10126-008-9124-7.
- Wu YH, Wu M, Wang CS, Wang XG, Yang JY, Oren A, Xu XW. 2008. Microbacterium profundi sp. nov., isolated from deep-sea sediment of polymetallic nodule environments. Int J Syst Evol Microbiol 58: 2930–2934. http://dx.doi.org/10.1099/ijs.0.2008/000455-0.
- Bentley DR, Balasubramanian S, Swerdlow HP, Smith GP, Milton J, Brown CG, Hall KP, Evers DJ, Barnes CL, Bignell HR, Boutell JM, Bryant J, Carter RJ, Keira Cheetham R, Cox AJ, Ellis DJ, Flatbush MR, Gormley NA, Humphray SJ, Irving LJ. 2008. Accurate whole human

- genome sequencing using reversible terminator chemistry. Nature **456**: 53–59. http://dx.doi.org/10.1038/nature07517.
- Li R, Li Y, Kristiansen K, Wang J. 2008. SOAP: short oligonucleotide alignment program. Bioinformatics 24:713–714. http://dx.doi.org/ 10.1093/bioinformatics/btn025.
- Li R, Zhu H, Ruan J, Qian W, Fang X, Shi Z, Li Y, Li S, Shan G, Kristiansen K, Li S, Yang H, Wang J, Wang J. 2010. *De novo* assembly of human genomes with massively parallel short read sequencing. Genome Res 20:265–272. http://dx.doi.org/10.1101/gr.097261.109.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25: 955–964. http://dx.doi.org/10.1093/nar/25.5.0955.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res 35:3100–3108. http://dx.doi.org/10.1093/ nar/gkm160.
- Griffiths-Jones S, Bateman A, Marshall M, Khanna A, Eddy SR. 2003.
  Rfam: an RNA family database. Nucleic Acids Res 31:439–441. http://dx.doi.org/10.1093/nar/gkg006.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.

- 11. Kanehisa M, Araki M, Goto S, Hattori M, Hirakawa M, Itoh M, Katayama T, Kawashima S, Okuda S, Tokimatsu T, Yamanishi Y. 2008. KEGG for linking genomes to life and the environment. Nucleic Acids Res 36:D480–D484. http://dx.doi.org/10.1093/nar/gkm882.
- Kanehisa M, Goto S. 2000. KEGG: Kyoto Encyclopedia of Genes and Genomes. Nucleic Acids Res 28:27–30. http://dx.doi.org/10.1093/nar/ 28.1.27.
- Moriya Y, Itoh M, Okuda S, Kanehisa M. 2005. KAAS: KEGG Automatic Annotation Server. Genome Informatics 5:2005. http://www.jsbi.org/pdfs/journal1/GIW05/GIW05P005.pdf.
- 14. Huston WM, Jennings MP, McEwan AG. 2002. The multicopper oxidase of *Pseudomonas aeruginosa* is a ferroxidase with a central role in iron acquisition. Mol Microbiol 45:1741–1750. http://dx.doi.org/10.1046/j.1365-2958.2002.03132.x.
- 15. Rensing C, Grass G. 2003. *Escherichia coli* mechanisms of copper homeostasis in a changing environment. FEMS Microbiol Rev 27:197–213. http://dx.doi.org/10.1016/S0168-6445(03)00049-4.
- 16. Singh SK, Grass G, Rensing C, Montfort WR. 2004. Cuprous oxidase activity of CueO from *Escherichia coli*. J Bacteriol 186:7815–7817. http://dx.doi.org/10.1128/JB.186.22.7815-7817.2004.
- 17. Grass G, Thakali K, Klebba PE, Thieme D, Müller A, Wildner GF, Rensing C. 2004. Linkage between catecholate siderophores and the multicopper oxidase CueO in *Escherichia coli*. J Bacteriol 186:5826–5833. http://dx.doi.org/10.1128/JB.186.17.5826-5833.2004.